

How to produce comparable data in conservation genetics for the Apennine brown bear

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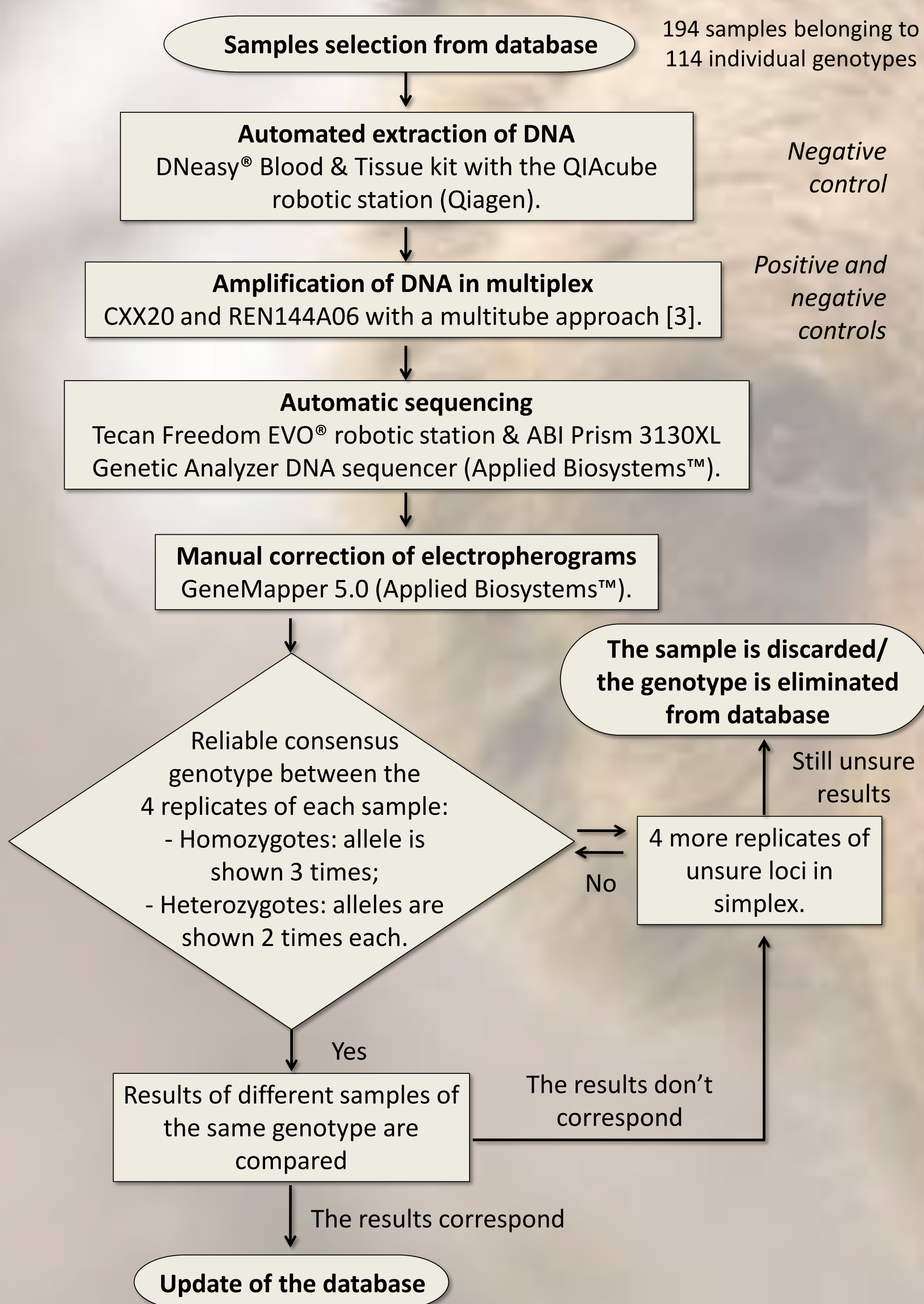
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INTRODUCTION

The Apennine brown bear (*U. arctos marsicanus*) presents a low level of variability [1], therefore ISPRA conducted the individual identification on the basis of 11 Ursidae specific markers (STR) plus sex. In the last decade, two different labs (WGI, Wildlife Genetics International, B.C., Canada and ISPRA) conducted the genotyping of the Apennine brown bear. WGI added two markers designed on the domestic dog genome (CXX20 and REN144A06) and removed two ones that had been previously used. Thus, their total selection was of 11 markers, 9 of which in common with ISPRA (G1D, G10B, G10C, G10L, Mu05, Mu11, Mu50, Mu51, Mu59), with an additional marker in common to both labs for equivocal cases (G10P) [2]. For a population with a low variability it is important to select the optimal STR marker set for individual identification, in order to allow the correct identification of the individuals overtime and to reduce genotyping errors.

MATERIALS AND METHODS



The following software were used for data analysis:

- GenAlEx 6.4 for allelic patterns, H_o , H_e , HWE, P_{ID} , $P_{ID_{Sib}}$, number of MM.
- GIMLET 1.3.3 and MicroChecker 2.2.3 to estimate genotyping errors frequencies (ADO, FA, PCR+ and null alleles).
- R (*chisq.test* and *fisher.test*) for statistical significance among groups.

CONCLUSIONS

- In order to avoid both underestimation (high values of P_{ID}) and overestimation (high levels of ADO and FA) in genotyping results, future monitoring will be conducted using the ISPRA set of 11 Ursidae-specific STRs with the addition of CXX20, that minimize the risk of shadow effect ($P_{ID} = 8.6 \cdot 10^{-6}$; $P_{ID_{Sib}} = 3.0 \cdot 10^{-3}$). In addition, marker REN144A06 will be used to improve the discriminatory capacity in uncertain cases.
- The population shows a slight and not significant loss of diversity due to genetic drift (Fig. 6, Fig. 7). Therefore the chosen STR panel is suitable for individual identification in the near future, but markers with higher discriminatory power will be needed for parentage analysis (eg. panel of SNPs).

RESULTS

The comparison of the different marker sets (Tab. 1, Fig. 1-5) shows an improvement, albeit not significant, of the discrimination capacity using the complete set of 13 STRs + AMG compared to the other STRs marker sets. However canid loci show a higher occurrence of genotyping errors. Allelic patterns (Fig. 7) show slight variation over time and PCA (Fig. 6) shows a substantial overlap of genetic diversity in the two considered periods.

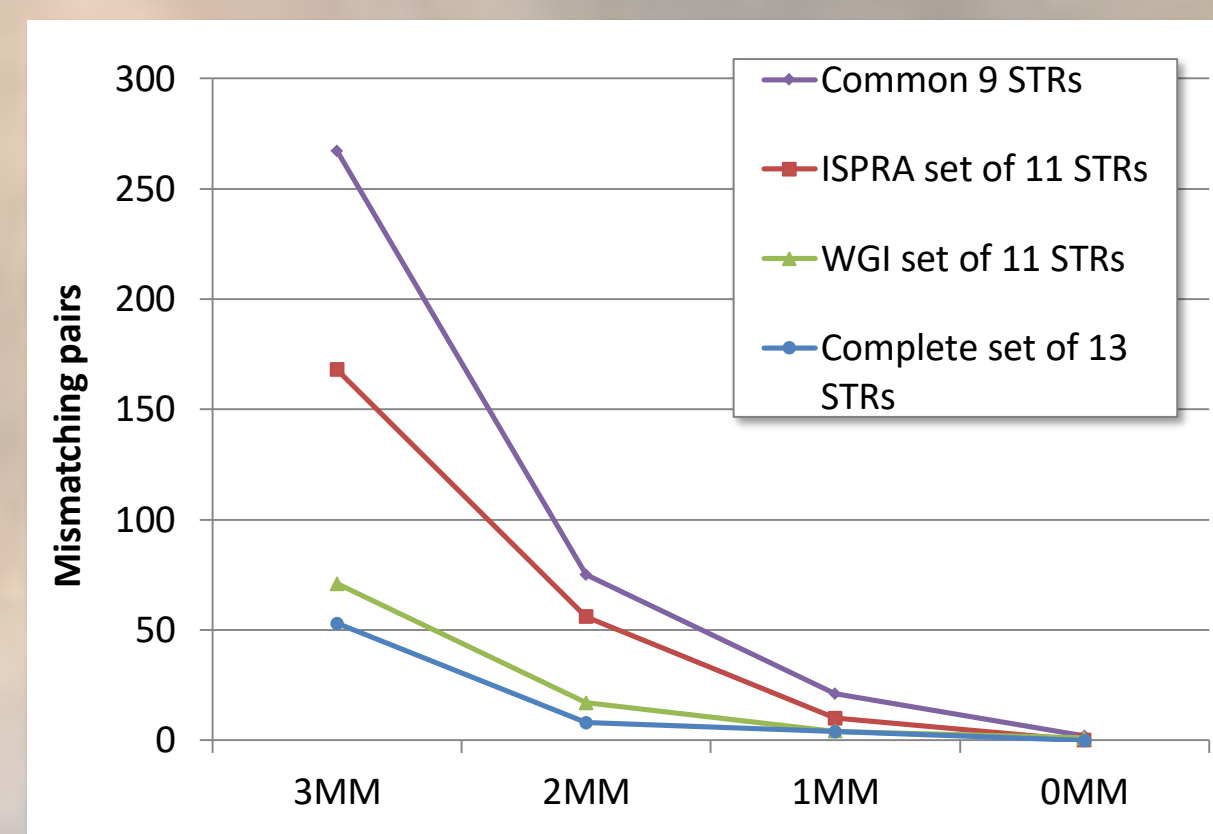


Fig. 1 - Number of mismatching pairs for each STR marker set.

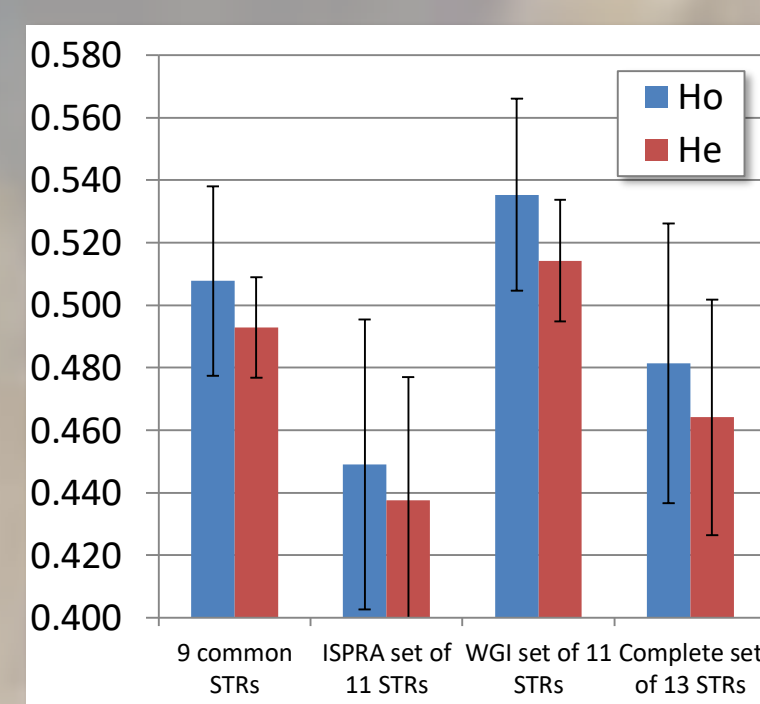


Fig. 3 - Mean H_o and H_e in the four STR marker sets.

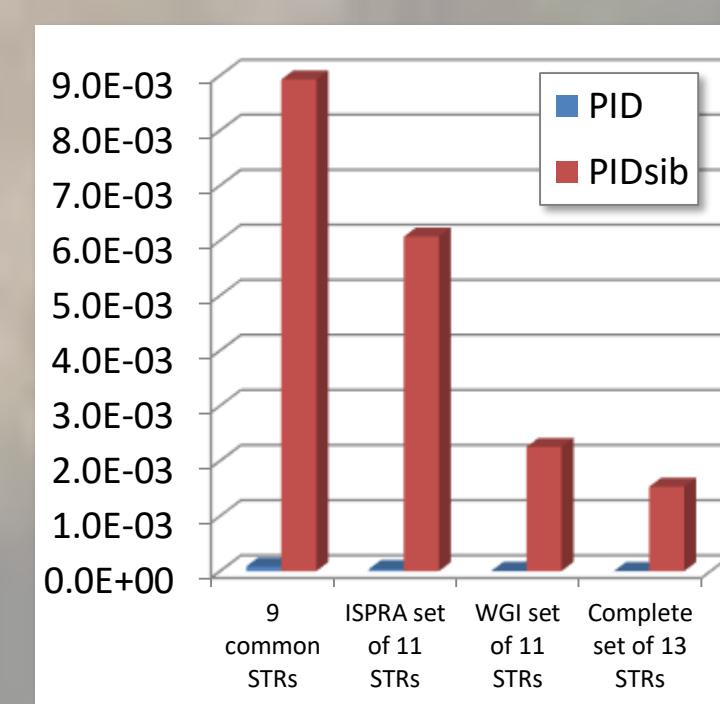


Fig. 4 - P_{ID} and $P_{ID_{Sib}}$ in the four STR marker sets.

Locus	Range (bp)	Conversion	P_{ID}	$P_{ID_{Sib}}$
CXX20	132-136	-3	0.22	0.50
REN144A06	110-130	+1	0.24	0.51
G1D	100-114	-72	0.25	0.53
Mu51	114-122	-92	0.26	0.53
G10B	112-128	-28	0.36	0.58
G10C	95-105	-102	0.37	0.59
Mu59	101-107	-128	0.38	0.60
Mu11	88-96	-100	0.39	0.62
Mu05	135-137	/	0.40	0.62
G10L	148-154	-9	0.41	0.63
Mu50	100-104	-32	0.41	0.63
G10P	152-164	+7	0.65	0.81
Mu15	117-121	Not used by WGI	0.71	0.84
Amelogenin	158-212	-46/-38	-	-

Table 1 - Range and conversion factors of the STRs.

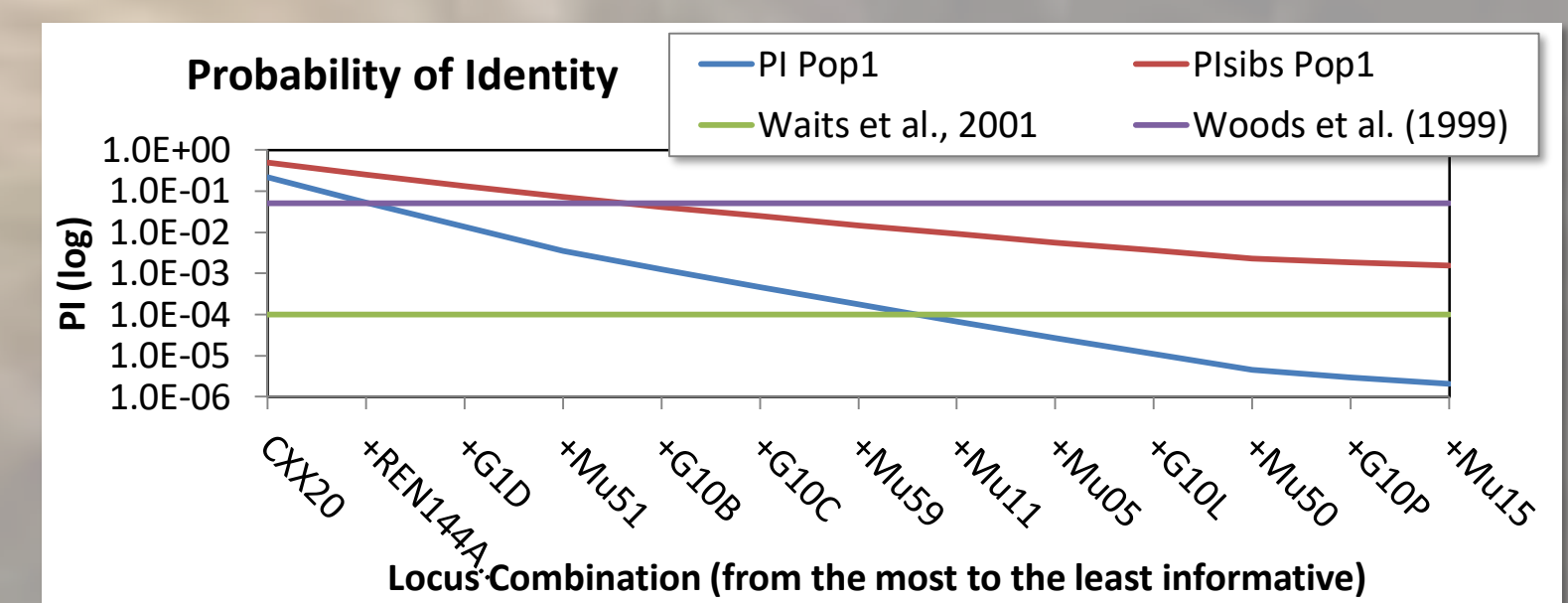


Fig. 2 - P_{ID} , $P_{ID_{Sib}}$, P_{ID} threshold of < 0.0001 (Waits *et al.*, 2001 [4]), $P_{ID_{Sib}}$ threshold of < 0.05 (Woods *et al.*, 1999 [5]).

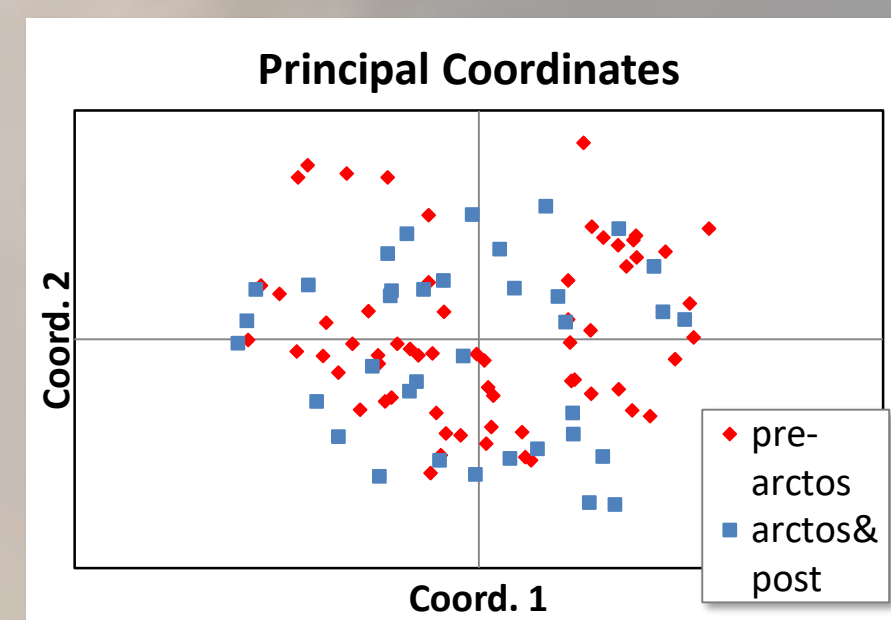


Fig. 6 - PCA of the variability from 2000-2010 to 2011-2017.

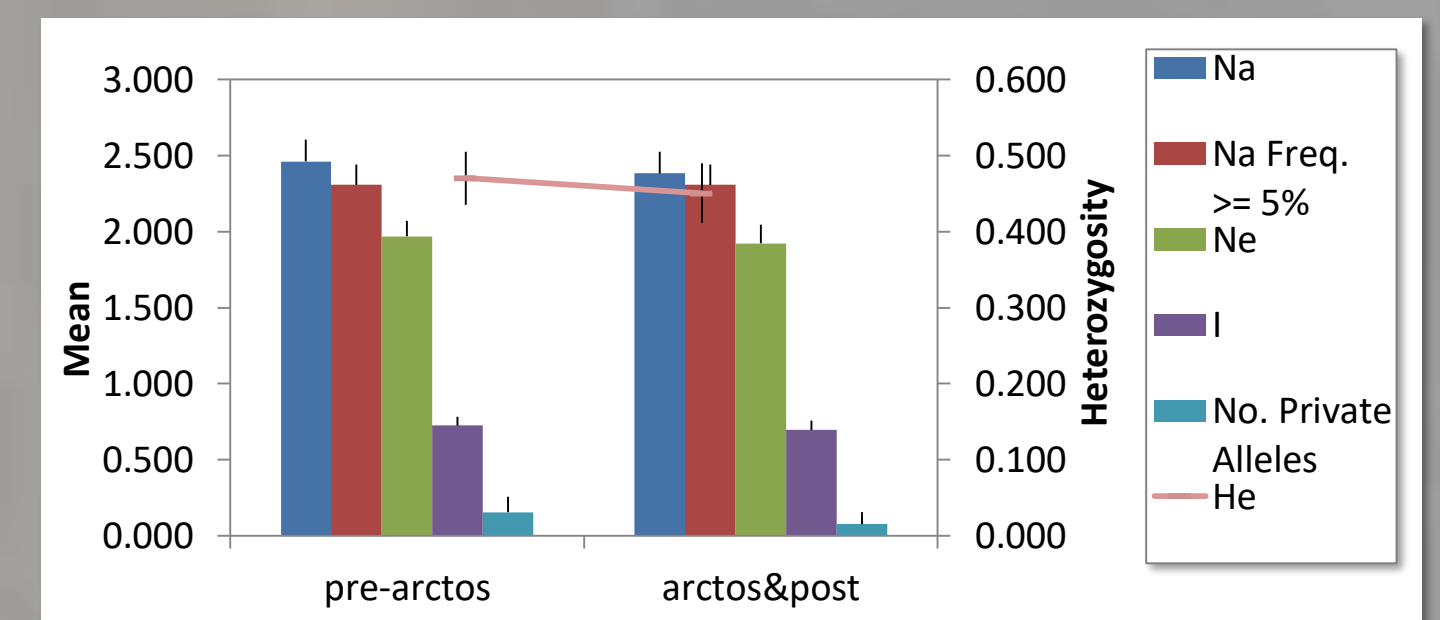


Fig. 7 - Variation of allelic patterns from 2000-2010 to 2011-2017.

ACKNOWLEDGMENTS

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